

## UgtP PROTEIN PURIFICATION

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### DAY 1:

- Streak out JC6 strain (*pBAD-ugtP*, *slyD*<sup>-</sup>) onto LB Amp plate.
- Prepare 4L autoclaved LB for induction (45min liquid cycle for every 2L)
  - 4L filter-sterilized ice-cold UgtP buffer with low salt concentration (10% glycerol, 50mM Tris pH8.0, 100mM NaCl) for dialysis
  - ~100ml filter-sterilized ice-cold UgtP buffer with high salt concentration (10% glycerol, 50mM Tris pH8.0, 500mM NaCl)
  - 4ml 100µg/ml Amp
  - 40ml 20% filter-sterilized L-Arabinose

### DAY 2:

- Grow JC6 from single colonies in 2X25 ml LB Amp O/N.

### DAY 3:

- Add 1ml Amp to each 1L autoclaved LB prior to induction.
- Inoculate 10ml O/N culture into each 1L LB Amp. At OD<sub>600</sub>~0.4 (~2-3hrs), induce each 1L culture with 10ml 20% L-Arabinose. Let grow for 4hrs at 37°C.
- Pellet the culture using the big rotor in the Kunkel centrifuge (always fill the rotor, balance, and never fill the bottle above the "line"): rotor code 3, 4000rpm, 10min.
- Resuspend each pellet in 10ml ice-cold UgtP buffer (high salt concentration: 500mM NaCl). Transfer to 50ml conical tubes and centrifuge again: 6000rpm, 15min, 4°C (fast cool before use). Pour off the supernatant and store pellets at -80°C or proceed to the next step.

### DAY 4:

- Resuspend 2 pellets in 30ml UgtP buffer with high salt concentration (10% glycerol, 50mM Tris pH 8.0, 500mM NaCl) with 300µl AEBSF.
- French Press 2-3 times.
- Transfer to 70Ti tubes. Balance to within 0.5g. Centrifuge in Ultra: 40Krpm, 45min, 4°C (70Ti rotor is kept in cold room. Cooling down Ultra takes ~1hr).
- Pre-run AKTA Prime and load the supernatant.

AKTA Prime Pre-run: 1) Run 20ml ddH<sub>2</sub>O from A2 to Waste at 20ml/min

- 2) Fill superloop with ddH<sub>2</sub>O
- 3) Inject 10ml ddH<sub>2</sub>O from superloop at 5ml/min
- 4) Run 20ml ddH<sub>2</sub>O to Load at 5ml/min
- 5) Run 20ml ddH<sub>2</sub>O from A1 to Waste at 20ml/min
- 6) Put UgtP buffer (high salt) in A1
- 7) Run 20ml ddH<sub>2</sub>O from A3 to Waste at 20ml/min
- 8) Put NiSO<sub>4</sub> in A3
- 9) Run 20ml ddH<sub>2</sub>O from A4 to Waste at 20ml/min
- 10) Put EDTA in A4
- 11) Run 20ml ddH<sub>2</sub>O from B to Waste at 20ml/min
- 12) Put UgtP buffer (high salt) with imidazole in B
- 13) Run 20ml of UgtP buffer from A1 to Waste at 20ml/min

- 14) Use a glass pipette to push superloop to 40ml
  - 15) Fill superloop with UgtP buffer (high salt)
  - 16) Inject 10ml UgtP buffer from superloop at 5ml/min
  - 17) Load 30ml sample. Put in collecting tubes.
  - 18) Turn on the lamp. Turn on the recorder.
  - 19) Run stored method 38.
  - 20) Fractions 6, 7, and 8 should contain the most protein
- Note: Pressure should always be kept at 0.3

- Dialyze fractions 6, 7, and 8 into UgtP buffer with low salt concentration (100mM NaCl) twice (once O/N, and once for another 2hrs) in the cold room using dialysis tubing, clamps (Qingwei's drawer), 1L beakers, stir bars and stir plates.

#### DAY 5:

- Concentrate dialyzed UgtP fractions using Centricon YM-30 until ~0.5ml is left in the Centricon: 4500rcf, 30min or more, 4°C. Flip the Centricon upside down to elute the protein by centrifuging again: 1000rcf, 10min, 4°C.
- Aliquot 25µl UgtP into small eppi tubes and dump into liquid nitrogen right away. Store the aliquots at -80°C.
- Clean-up AKTA Prime.

AKTA Prime Clean-up: 1) Open superloop and rinse it well

- 2) Use a glass pipette to push superloop to 30ml
- 3) Fill the bottom of superloop with UgtP buffer (high salt)
- 4) Fill the top of superloop with ddH<sub>2</sub>O
- 5) Inject 5ml UgtP buffer from superloop at 5ml/min
- 6) Replace UgtP buffer with ddH<sub>2</sub>O in superloop
- 7) Inject 5ml ddH<sub>2</sub>O from superloop at 5ml/min
- 8) Run 20ml EDTA from A4 to Waste at 20ml/min
- 9) Run 40ml EDTA from A4 to Load at 5ml/min
- 10) Run 40ml ddH<sub>2</sub>O from A2 to Load at 5ml/min
- 11) Run 20ml ddH<sub>2</sub>O (flask) from A1, B, A4 and A3 to Waste at 20ml/min
- 12) Put filter lines into EtOH flask (large EtOH flask for A4)
- 13) Run 20ml EtOH from A4 to Waste at 20ml/min
- 14) Empty superloop and fill with 20% EtOH
- 15) Inject 5ml EtOH from superloop at 3ml/min
- 16) Run 20ml EtOH from A4 to Load at 3ml/min
- 17) Replace large EtOH flask in A4 with a small one
- 18) Run 20ml EtOH from B, A1, A2 and A3 to Waste at 20ml/min
- 19) Turn off the machine. Refill ddH<sub>2</sub>O flask with fresh.