

Convergent Evidence for Predispositional Effects of Brain Gray Matter Volume on Alcohol Consumption

David A.A. Baranger, Catherine H. Demers, Nourhan M. Elsayed, Annchen R. Knodt, Spenser R. Radtke, Aline Desmarais, Lauren R. Few, Arpana Agrawal, Andrew C. Heath, Deanna M. Barch, Lindsay M. Squeglia, Douglas E. Williamson, Ahmad R. Hariri, and Ryan Bogdan

ABSTRACT

BACKGROUND: Alcohol use has been reliably associated with smaller subcortical and cortical regional gray matter volumes (GMVs). Whether these associations reflect shared predisposing risk factors or causal consequences of alcohol use remains poorly understood.

METHODS: Data came from 3 neuroimaging samples ($N = 2423$), spanning childhood or adolescence to middle age, with prospective or family-based data. First, we identified replicable GMV correlates of alcohol use. Next, we used family-based and longitudinal data to test whether these associations may plausibly reflect a predispositional liability for alcohol use or a causal consequence of alcohol use. Finally, we used heritability, gene-set enrichment, and transcriptome-wide association study approaches to evaluate whether genome-wide association study-defined genomic risk for alcohol consumption is enriched for genes that are preferentially expressed in regions that were identified in our neuroimaging analyses.

RESULTS: Smaller right dorsolateral prefrontal cortex (DLPFC) (i.e., middle and superior frontal gyri) and insula GMVs were associated with increased alcohol use across samples. Family-based and prospective longitudinal data suggest that these associations are genetically conferred and that DLPFC GMV prospectively predicts future use and initiation. Genomic risk for alcohol use was enriched in gene sets that were preferentially expressed in the DLPFC and was associated with replicable differential gene expression in the DLPFC.

CONCLUSIONS: These data suggest that smaller DLPFC and insula GMV plausibly represent genetically conferred predispositional risk factors for, as opposed to consequences of, alcohol use. DLPFC and insula GMV represent promising biomarkers for alcohol-consumption liability and related psychiatric and behavioral phenotypes.

Keywords: Alcohol, Gene expression, Heritability, Imaging, Longitudinal, Structure

<https://doi.org/10.1016/j.biopsych.2019.08.029>

Alcohol use and its associated negative consequences are ubiquitous international public health concerns. Worldwide, the average person 15 years of age or older consumes 6.2 liters of alcohol annually, and alcohol use accounts for 6% of deaths and 5% of disease burden (1). Combined with the widespread prevalence of problematic alcohol use [e.g., alcohol use disorder lifetime prevalence = 29% (2); current-month binge drinking = 26% of adults in the United States (3)], these staggering public health consequences have led to extensive efforts to understand the impact of alcohol use on brain and behavior and to identify markers of alcohol use liability.

Neuroimaging studies have consistently shown that alcohol consumption and alcohol use disorder are associated with smaller subcortical and cortical gray matter volumes (GMVs), particularly among regions that feature prominently in emotion, memory, reward, cognitive control, and decision making (4–10). While there is evidence that these associations may

arise as a consequence of drinking (e.g., reduced neurogenesis in nonhuman primate models, greater GMV decline among adolescents following the initiation of heavy drinking, GMV normalization following abstinence from alcohol among alcohol-dependent individuals) (9,11–18), other data suggest that such associations may reflect preexisting vulnerabilities that precede and predict drinking initiation and escalating use (19–23).

Here, using neuroimaging data from 3 samples ($N = 2423$) (24–26) spanning childhood and adolescence to middle age with prospective or family-based data, we first identified replicable GMV correlates of alcohol use before testing whether these correlates 1) are plausibly attributable to shared predisposing factors (e.g., shared genetic influence) or arise as a consequence of alcohol use, 2) prospectively predict future drinking in young adulthood, and 3) predict drinking initiation in adolescence. Finally, using curated postmortem data, we

examined whether genetic risk for alcohol consumption is associated with genes and genetically conferred differences in gene expression that are preferentially expressed in regions identified by neuroimaging analyses or the brain more generally. Here, we applied gene-set enrichment, partitioned heritability, and transcriptome-wide association study (TWAS) (27) analyses to genome-wide association study (GWAS) summary statistics from the UK Biobank ($N = 112,117$) (28) and Alcohol Genome-Wide Consortium and the Cohorts for Heart and Aging Research in Genomic Epidemiology Plus consortia (AlcGen/CHARGE+) ($N = 70,460$) (29) studies of alcohol consumption, and RNA-seq data from the Genotype-Tissue Expression (GTEx) project ($n = 81\text{--}103$) (30) and the Common Mind Consortium ($N = 452$) (31).

METHODS AND MATERIALS

Participants

Neuroimaging data were drawn from 3 independent studies—the Duke Neurogenetics Study (DNS) ($n = 1303$) (26), the Human Connectome Project (HCP) ($n = 897$) (24), and the Teen Alcohol Outcomes Study (TAOS) ($n = 223$) (25)—that assessed behavioral, experiential, and biological phenotypes among young adult college students (DNS sample), young to middle-aged adults (HCP sample), and children and adolescents (TAOS sample). The DNS and TAOS studies collected longitudinal data on alcohol use subsequent to the baseline scan. The family-based HCP sample is composed of twin and non-twin siblings. All studies followed protocols approved by relevant institutional review boards and remunerated participants. Additional information regarding each sample is provided in Supplement 1.

Alcohol Use Assessment

Alcohol use in the DNS was assessed at baseline (past 12-month use) and follow-ups (questions modified to reflect use following the prior assessment) using the Alcohol Use Disorders Identification Test consumption subscale (AUDIT-C) (DNS: $\alpha = .85$; mean = 3.76; SD = 2.64; range = 0–12) (32,33). The AUDIT-C was approximated (aAUDIT-C) in the HCP sample ($\alpha = .786$; mean = 3.42; SD = 2.65; range = 0–12) and TAOS ($\alpha = .893$; mean = 0.45; SD = 1.26; range = 0–9) using questions from Semi-Structured Assessment for the Genetics of Alcoholism (34) and Substance Use Questionnaire (35), respectively. In TAOS, the initiation of alcohol use was defined as attaining a score of ≥ 1 on the aAUDIT-C (i.e., participant reports consuming ≥ 1 full alcoholic beverage; $n = 82$ started during the study; age (in years): mean = 16.68, SD = 1.39, range 14.12–19.64). Supplement 1 contains additional details.

Covariates

Variables known to be correlated with alcohol consumption, GMV, or both were included as covariates in all analyses: age (36–38), sex (37–40), ethnicity (40,41), socioeconomic status (SES) (36–40), early-life and recent-life stress (42–44), and intracranial volume. In adult samples (DNS, HCP), the presence of any nonsubstance Axis I DSM-IV psychiatric disorder was included as a covariate. As the TAOS sample was composed of children and adolescents enriched for a family history of

depression, Tanner stage and depressive symptoms were included as covariates. Supplement 1 contains additional details, including consideration of nicotine and cannabis use.

Magnetic Resonance Imaging Processing

Acquisition parameters and GMV processing for each study are described in Supplement 1.

Statistical Analyses

Discovery—DNS. A whole-brain voxel-based morphometry generalized linear model regression analysis was conducted using SPM12 to test whether alcohol consumption (AUDIT-C) is associated with differences in GMV. Covariates included sex, age, self-reported race/ethnicity (i.e., not-white/white, not-black/black, not Hispanic/Hispanic), scanner identification (2 identical scanners were used), intracranial volume, presence of a diagnosis other than alcohol or substance abuse or dependence, perceived stress, parental education level, early-life stress (assessed via the Childhood Trauma Questionnaire), and perceived SES. Analyses were thresholded at $p < .05$ familywise error corrected with a cluster extent threshold of 10 contiguous voxels ($k_e = 10$) across the entire search volume.

Replication—HCP. Analyses examined whether alcohol consumption (aAUDIT-C) predicted GMV only within regions of interest (ROIs) where associations were observed in the discovery DNS sample (Figure 1, Table S1 in Supplement 1). ROIs were defined by the Automated Anatomic Labeling atlas (45). A voxelwise generalized linear model regression was conducted using multilevel block permutation-based nonparametric testing (FSL PALM v.alpha103; tail approximation $p < .10$ with 5000 permutations), which accounts for the family structure of the HCP data while correcting for multiple comparisons (46–48). Covariates included sex, age, self-reported race and/or ethnicity, intracranial volume, twin and/or sibling status (dizygotic or not, monozygotic or not, half-sibling or not), presence of a diagnosis other than alcohol or substance abuse or dependence, perceived stress, education level, and SES. Analyses were thresholded at $p < .05$ familywise error corrected with a cluster extent threshold of 10 contiguous voxels ($k_e = 10$).

Post Hoc Analyses

Total anatomical GMV of ROIs associated with alcohol use in both the DNS and HCP (i.e., right insula and middle and superior frontal gyri) (see Results) were extracted from both datasets for post hoc analyses. Total volumes were used so that effect sizes would not be inflated by selecting only voxels that were specifically associated with the outcome of interest (49).

Heritability. SOLAR-Eclipse software (<http://solar-eclipse-genetics.org>) (50), in conjunction with the R package solarius (51), which uses maximum likelihood variance decomposition methods, was used to estimate phenotypic heritability (h^2 , the fraction of phenotypic variance attributable to additive genetic factors), as well as genetic (ρ_g) and unique environmental (ρ_e) correlations (i.e., the fraction of the correlation between 2 phenotypes that is attributable to either additive genetic or

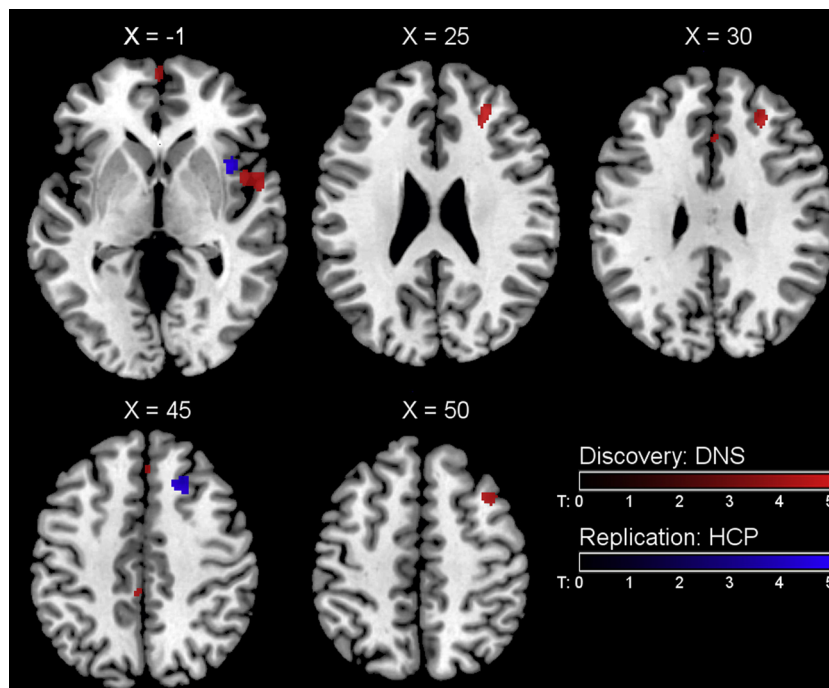


Figure 1. Identification of replicable volumetric associations with alcohol consumption. This statistical parametric map illustrates regions of reduced brain volume associated with increased alcohol consumption (Table S1 in Supplement 1), which are overlaid onto a canonical structural brain image Montreal Neurological Institute coordinates and statistics (Duke Neurogenetics Study [DNS]: $p < .05$, familywise error whole-brain corrected, ≥ 10 contiguous voxels; Human Connectome Project [HCP]: $p < .05$, familywise error region-of-interest corrected, ≥ 10 contiguous voxels). Alcohol consumption was not associated with increased volume in any region. Notably, in the HCP dataset, the superior frontal gyrus cluster extended into the right middle frontal gyrus and was located relatively far (34 mm dorsal) from the original right superior frontal cluster identified in the DNS. In contrast, this peak in the HCP was located 11.6 mm away from the right middle frontal peak identified in the DNS. Thus, for the purposes of post hoc analyses, the combined volume of both the right middle and superior frontal gyri cortices was extracted from both samples. Cluster overlap at an uncorrected threshold and comparison of effect sizes are shown in Figures S2 and S3 in Supplement 1.

individual-specific environmental factors, respectively) of GMV and alcohol consumption. Post hoc analyses (Supplement 1) assessed the contribution of shared environmental factors to phenotypic correlations (ρ_c). These analyses were conducted among the subset of related participants from the HCP ($n = 804$; 293 families, 115 monozygotic and 64 dizygotic twin pairs and 422 nontwin siblings, excluding singletons and half-siblings). Covariates were identical to those in neuroimaging analyses. To ensure normality of measurements and accuracy of estimated parameters, an inverse normal transformation was applied to all continuous traits and covariates prior to analyses.

Discordant Twin and Nontwin Sibling Analysis. Following evidence that alcohol consumption is heritable with volume of the right insula and middle and superior frontal gyri (see Results), we examined whether same-sex twin and nontwin sibling pairs discordant for alcohol consumption differed from each other on brain volume in the HCP sample. These analyses examined whether aAUDIT-C was associated with insular or middle and superior frontal volume after accounting for sibling-shared genetic background and experience. Same-sex siblings were considered “high alcohol consumers” if their aAUDIT-C score was >0.5 SD above the sample mean (aAUDIT-C > 4.67), or “low alcohol consumers” if their score was <0.5 SD below the sample mean (aAUDIT-C < 1.54), respectively. A concordant sibling pair was defined as a pair who were both in the same category of consumption (i.e., high or low) and additionally scored within 1 SD of each other (low alcohol concordant pairs: $n = 117$; aAUDIT-C mean = 0.84, SD = 0.77; high alcohol concordant pairs: $n = 54$; aAUDIT-C mean = 7.08, SD = 1.4). There were 72 discordant sibling pairs (“low discordant”; aAUDIT-C mean = 1.25, SD = 0.73; “high discordant”; aAUDIT-C mean = 6.47, SD = 1.67).

Participants could be included in >1 pair ($n = 368$ individuals) when considering relationships with multiple siblings. Discordancy analyses were conducted using linear mixed models, using the psych (52) and lme4 (53) packages in R (54) to account for the multiple-sibling structure within families. Covariates were identical to those used in neuroimaging analyses. Additional information on models tested and their interpretation are available in Supplement 1.

DNS Longitudinal Changes in Alcohol Consumption. Hierarchical density-based clustering (R dbSCAN package) (55) was used to detect and remove temporal outlier responses to the follow-up questionnaire (Supplement 1, Figure S5 in Supplement 1). The R nlme package (56) was used to fit a longitudinal multilevel linear model to examine whether GMV predicted AUDIT-C at follow-ups. The model included both random intercept and random slope components with a continuous autoregressive correlation structure. Time was coded as both linear and quadratic age at the date of response (baseline or follow-up). Models tested the interaction between brain volume and age (i.e., does baseline ROI volume predict a different slope of change in drinking behavior as participants age?). Covariates were z-scored, and they were identical to those used in neuroimaging analyses, with the addition of second-order interactions between covariates and primary variables (57,58). Each of the 2 ROIs was tested in a separate model, and p values were false discovery rate (FDR) corrected (i.e., 4 tests: middle \times linear age, superior \times linear age, middle \times quadratic age, superior \times quadratic age).

TAOS Longitudinal Initiation of Alcohol Use. The R lme4 package (59) was used to fit a longitudinal logistic

multilevel model, which tested whether baseline brain volume in nondrinking adolescents predicted future initiation of alcohol use. The model included both random intercept and random slope components, and time was coded as both the linear and quadratic age at the date of response. The model tested the interaction between GMV and age (i.e., does baseline ROI volume predict a different likelihood of initiation as participant's age?). Covariates were z-scored, and they included demographic variables (age, sex, ethnicity, and SES), stress (Childhood Trauma Questionnaire and Stressful Life Events Schedule), Tanner stage, Mood and Feelings Questionnaire scores, family history of depression, age at magnetic resonance imaging scan, and intracranial volume. Second-order interactions between covariates and primary variables (e.g., middle frontal volume \times sex, middle superior volume \times SES, age \times sex, age \times SES) were also included (58). Each of 2 ROIs, right superior frontal cortex and right middle frontal cortex, was tested in a separate model. Insula volume was excluded, as it was not significant in DNS longitudinal analyses. The p values were subsequently FDR corrected (4 tests).

Single Nucleotide Polymorphism-Based Enrichment. We tested whether the single nucleotide polymorphism (SNP)-based heritability of alcohol consumption is enriched in brain-expressed gene sets and whether this enrichment is specific to any region. Stratified linkage disequilibrium-score regression (60–62) was applied to summary statistics from the GWAS of alcohol consumption in the UK Biobank ($N = 112,117$) (28). Tissue-enriched gene sets were generated using data from the GTEx Consortium (30,61). A gene is assigned to a gene set if it shows greater enrichment in that tissue than 90% of genes. It was further tested whether genetic associations with alcohol consumption are enriched in brain-expressed gene sets using the analysis tool MAGMA (63) implemented through the platform FUMA (64).

Transcriptome-wide Analysis. We tested whether genetic risk for alcohol consumption is predictive of

differences in postmortem gene expression. Precomputed gene-expression RNA-sequencing weights for 9 brain regions and the liver from the GTEx project (30) were analyzed using the FUSION suite (27). Analyses used GWAS results for alcohol consumption from the UK Biobank (28). Results were Bonferroni-corrected for $n = 9839$ tests across the 10 tissues (Supplemental Data). Replication was tested using an independent alcohol-consumption GWAS ($N = 70,460$) (29) and dorsolateral prefrontal cortex (DLPFC) gene-expression weights from the CommonMind Consortium (31). As the gene that showed the strongest association in the discovery dataset was not present in the replication data, we examined whether any of the Brodmann area 9 (BA 9) gene-expression associations at FDR-corrected $p < .05$ were significant in the replication data (see Results). Replicated genes were probed for association with other GWAS phenotypes using a phenome-wide association study (PheWAS) implemented through the “GWAS Atlas” browser (65). BA 9 in the GTEx dataset and DLPFC in the CommonMind consortium dataset overlap with the prefrontal regions implicated in our neuroimaging analyses (i.e., middle and superior frontal gyri) (see Results). No postmortem insula data were available.

RESULTS

Whole-brain discovery analyses in DNS revealed that greater alcohol consumption is associated with lower GMV across 8 clusters (Figure 1, Table 1) that encompass regions identified in prior studies of unselected samples (5,17) and among individuals with alcohol use disorder (4,6). The associations with 2 of these clusters (right insula, right middle and superior frontal gyri) replicated within an ROI analysis in the HCP (Figure 1, Table 1). Post hoc analyses revealed that effect sizes were nearly identical in the 2 samples, that results were equivalent when excluding nondrinkers (Figure S2 in Supplement 1), and that associations between alcohol use and GMV remained largely unchanged and significant when correcting for tobacco

Table 1. Location of Volumetric Reductions Associated With Alcohol Consumption

Index	No. of Voxels	p , Familywise Error Corrected	t	x, mm ^a	y, mm ^a	z, mm ^a	AAL-Atlas Location
Duke Neurogenetics Study							
1	279	.003	5.24	27	39	25	R middle frontal
1.b		.010	4.95	32	50	23	R middle frontal
2	344	.004	5.14	56	3	0	R superior temporal
2.b		.006	5.05	48	6	−5	R insula
3	44	.005	5.08	0	63	−3	L medial orbital frontal
4	76	.007	5.03	38	18	51	R middle frontal
5	64	.007	5.03	−3	−33	42	L middle cingulum
6	23	.019	4.80	2	27	30	R middle cingulum
7	17	.023	4.75	29	62	8	R superior frontal
8	12	.029	4.59	2	33	45	R medial superior frontal
Human Connectome Project							
1	42	.003	4.92	38	12	0	R insula
2	88	.008	4.70	20	24	42	R superior/middle frontal

AAL, Automated Anatomical Labeling; L, left; R, right.

^aCoordinates are provided in Montreal Neurological Institute space.

Alcohol Consumption and Brain Volume

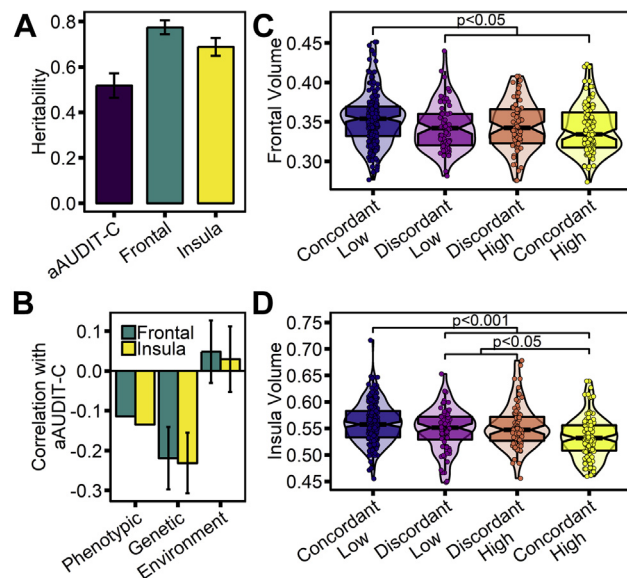


Figure 2. Shared genetic predisposition between alcohol consumption and brain volume. In the Human Connectome Project (HCP) sample, **(A)** alcohol consumption scores (approximated Alcohol Use Disorders Identification Test consumption subscale scores [aAUDIT-C]) and gray matter volume of the right insula and right middle and superior frontal cortices were all observed to be heritable (aAUDIT-C: 51.79%, $p < 2.2 \times 10^{-16}$; insula: 68.83%, $p < 2.2 \times 10^{-16}$; frontal: 74.46%, $p < 2.2 \times 10^{-16}$) (Table S1 in Supplement 1). **(B)** Significant phenotypic correlations between aAUDIT-C scores and volumes of the right insula and middle and superior frontal gyri are attributable to shared genetic factors (insula: -0.2314 , $p = .0022$; frontal: -0.2192 , $p = .0054$) but not unique environmental factors (Table S1 in Supplement 1). **(C)** Distribution of **(C)** right insula and **(D)** right middle and superior frontal volumes by alcohol exposure group. High = aAUDIT-C score $>$ sample mean $+ 0.5$ SD (i.e., > 4.67); Low = aAUDIT-C score $<$ sample mean $- 0.5$ SD (i.e., < 1.54); Concordant = both siblings are in the same alcohol exposure group; Discordant = one sibling is in the high group, while the other is in the low group. Contrast comparisons found evidence for predispositional effects of brain volume on alcohol consumption in both cases (insula: graded liability: $\beta = -0.0037$, $p = .049$, predispositional: $\beta = 0.0037$, $p = .0006$; frontal: predispositional: $\beta = 0.0019$, $p = .029$) (Table S2 in Supplement 1).

and cannabis use (Table S11 in Supplement 1). Statistics are presented in the table and figure legends.

Family-based analyses in the HCP ($n = 804$) revealed that alcohol consumption and GMV of the right insula and right middle/superior frontal gyrus are moderately to largely heritable (Figure 2A; Table S1 in Supplement 1). Moreover, decomposition analyses showed that phenotypic correlations between frontal and insular GMV and alcohol consumption are attributable to shared genetic, but not unique environmental, influences (Figure 2B, Table S1 in Supplement 1). Post hoc analyses confirmed that shared environmental factors did not significantly contribute to the correlation of alcohol consumption and GMV (Table S1 in Supplement 1). Analyses within twin and sibling pairs in the HCP sample who were concordant or discordant for alcohol use revealed that relative to siblings who were concordant for low alcohol use, siblings who were concordant for high use or discordant for use (i.e., 1 high use, 1 low use) had lower insular and frontal GMVs (Figure 2C, D; Table S2 in Supplement 1). Further, GMVs did not differ between low and high alcohol-using members of discordant

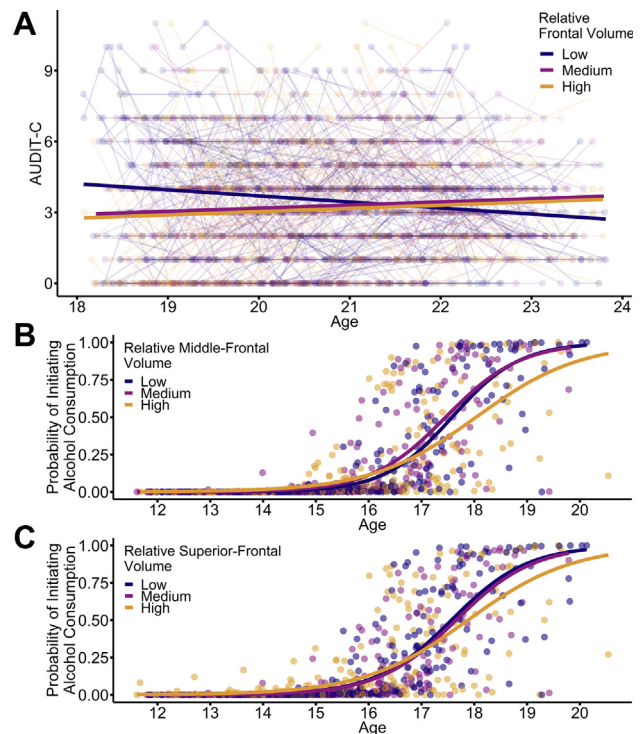


Figure 3. Frontal volume prospectively predicts alcohol use and initiation of consumption. **(A)** In the Duke Neurogenetics Study, participants with reduced volume of the right middle and superior frontal cortices reported elevated alcohol consumption before 20.85 years of age following the neuroimaging scan, and after accounting for baseline drinking (frontal \times age interaction: $\beta = 0.150$, false discovery rate–corrected $p = .008$) (Table S3 in Supplement 1). **(B, C)** In the Teen Alcohol Outcomes Study, participants with increased volume of the right middle and superior frontal cortices report initiation of alcohol consumption at an older age (midfrontal \times age interaction: $\beta = -57.042$, false discovery rate–corrected $p = .036$; superior frontal \times age interaction: $\beta = -60.74$, false discovery rate–corrected $p = .036$) (Table S4 in Supplement 1). Analyses were conducted with continuous data; the partition into 3 equally sized groups according to volume was done for display purposes only. AUDIT-C, Alcohol Use Disorders Identification Test consumption subscale.

pairs. As shared genetic and familial factors are matched within pairs, this pattern of results suggests that smaller frontal gyri and insula GMVs may reflect preexisting vulnerability factors associated with alcohol use, as opposed to a consequence of alcohol use.

Using available longitudinal data from the DNS ($n = 674$), lower GMV of the right frontal gyri, but not insula, predicted increased future alcohol consumption, over and above baseline consumption, but only in individuals who are under the legal age of drinking (i.e., younger than 21 years of age) in the United States (Figure 3A, Table S3 in Supplement 1). Similarly, in the TAOS longitudinal sample of children and adolescents, lower right middle and superior frontal gyri GMV predicted the initiation of alcohol use at an earlier age in those who were nondrinkers at baseline (Figure 3B, C; Table S4 in Supplement 1).

Gene-based association and partitioned heritability enrichment analyses of the UK Biobank GWAS of alcohol consumption revealed enrichment only among brain gene sets

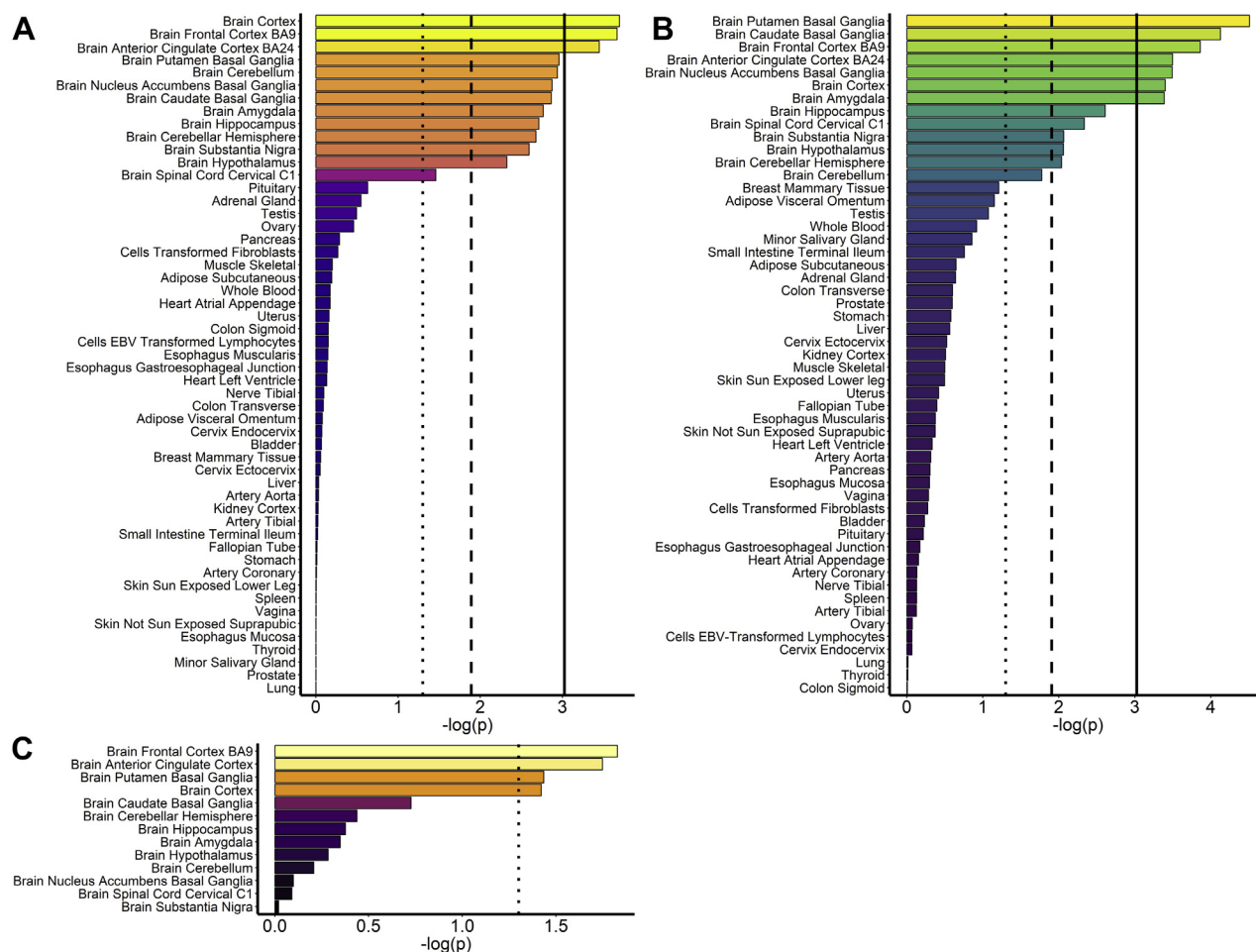


Figure 4. Tissue-specific enrichment of alcohol-consumption genomic risk. Enrichment of alcohol-consumption genome-wide association study (UK Biobank, $N = 112,117$) (A) associations and (B, C) heritability, in gene sets defined by the relative expression of genes (A, B) across all tissues and (C) within the brain, in the Genotype-Tissue Expression project dataset (Supplemental Data). The x-axis and color scale represent the significance of the enrichment (negative logarithmic scale of the p value). Solid, dashed, and dotted lines represent Bonferroni-corrected, false discovery rate-corrected, and nominally significant p values, respectively. BA, Brodmann area; EBV, Epstein-Barr virus.

(Figure 4). Moreover, BA 9, which overlaps with the frontal region identified in neuroimaging analyses, was among the regions with strongest enrichment (Figure S4 in Supplement 1, Supplemental Data). A TWAS analysis of these GWAS data similarly found that genetic risk for alcohol consumption was significantly associated with differences in gene expression across the brain within the GTEx dataset, including expression of *C16orf93* within BA 9 (Figure 5, Table 2, Supplemental Data). *C16orf93* was not available in the TWAS replication dataset [i.e., the dataset of Schumann *et al.* (29) and the CommonMind Consortium (31)] (Table 2). Three additional genes survived FDR correction in BA 9, two of which (i.e., *CWF19L1* and *C18orf8*) were available in the TWAS replication dataset (Figure 5, Table 2)¹. Genomic risk for alcohol consumption was significantly predictive of differential expression of *CWF19L1*

and *C18orf8* within the DLPFC of our TWAS replication dataset, in the same direction as was observed in the discovery dataset (Table 2, Supplemental Data). Notably, genetic risk for alcohol consumption was not significantly associated with the expression of any gene in the liver (Figure 5). A phenome-wide association study using the GWAS Atlas revealed evidence that both *CWF19L1* and *C18orf8* have been implicated in a host of phenotypes, including psychiatric conditions and related traits such as executive function and schizophrenia (*CWF19L1*), and substance use (*C18orf8*) (Supplemental Data).

Behavioral variables that might mediate links between brain structure and alcohol consumption (i.e., IQ, delay discounting, self-reported impulsivity, negative urgency, and neuroticism) and that were available in the DNS and HCP data sets and were tested for association with GMV. Despite nominally significant associations in the DNS between delay discounting and IQ and GMV of the right frontal cortex, none of these associations replicated within the HCP data or were robust to

¹Notably, correcting for only tests within BA 9 based on our neuroimaging results, *CWF19L1* remains significant following Bonferroni correction.

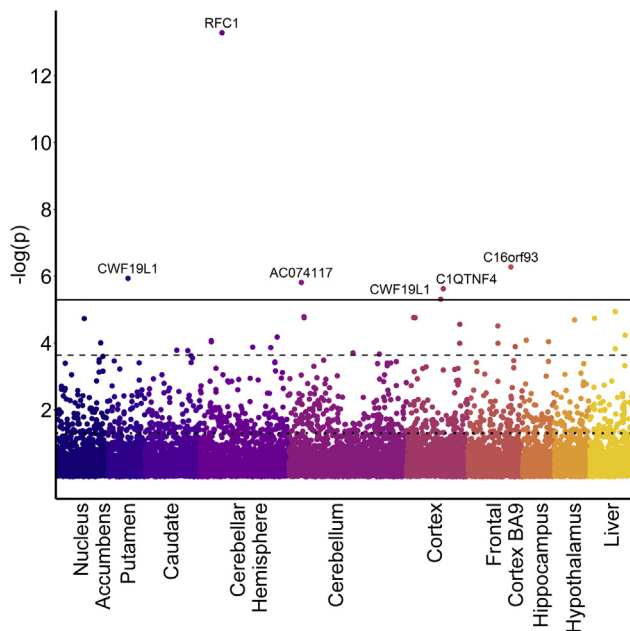


Figure 5. Transcriptome-wide association study of alcohol consumption predicting gene expression. Genetic risk for alcohol consumption according to the UK Biobank genome-wide association study ($n = 112,117$) is associated with differences in human postmortem gene expression (Genotype-Tissue Expression project; $n_s = 81-103$), including frontal cortex Brodmann area (BA) 9 (Supplemental Data). Notably, associations in the liver (far-right panel) do not survive Bonferroni correction for multiple comparisons, though 4 are significant at a less-stringent false discovery rate-based correction. The y-axis represents the significance of the association. Solid, dashed, and dotted lines represent Bonferroni-corrected, false discovery rate-corrected, and nominally significant p values, respectively.

multiple testing correction (see Supplement 1, Tables S9, S10 in Supplement 1).

DISCUSSION

We report convergent evidence that smaller GMVs of the right insula and DLPFC (i.e., middle and superior frontal gyri) plausibly represent genetically conferred liabilities that promote early alcohol use. First, we show that smaller GMVs of the right insula and DLPFC were replicably associated with alcohol use in 2 large neuroimaging samples. Second, family-based data

provide evidence that these associations are attributable to shared genetic factors with no evidence of a causal association, or that shared or unique environmental factors contribute to this association. Third, reduced DLPFC volume prospectively predicted future alcohol use among young adults as well as alcohol use initiation during adolescence among children and adolescents who were unexposed to alcohol at baseline. Finally, we found evidence that genomic risk for alcohol use is enriched among genes that are preferentially expressed within the DLPFC and is replicably predictive of gene expression in the DLPFC. Collectively, these convergent data suggest that lower GMVs in the middle and superior frontal gyri and insula may represent a preexisting genetic liability for drinking that could serve as a prognostic biomarker. Further, these data suggest that the alcohol use in the general population does not induce reductions in GMV, at least as measured using magnetic resonance imaging, as has been previously hypothesized (5,7,9). It is possible that reduced GMVs in the middle and superior frontal gyri and insula may promote alcohol use, increasing the likelihood of heavy use, which may then further potentiate GMV loss in these regions and others (9,11,12).

A few notable points within our data require additional interpretation. In the longitudinal child and adolescent sample of baseline nonusers (i.e., the TAOS sample), we found that DLPFC GMV prospectively predicts an early age of drinking initiation. In the DNS longitudinal prospective data of young adults, reduced GMV in these regions also predicted future alcohol use, even after accounting for the extent of baseline alcohol use. However, this characteristic was only predictive up until 20.85 years of age. It is possible that risk conferred by reduced GMV in the DLPFC is developmentally constrained or may be minimized by environmental differences in permissivity or legality, as the legal drinking age in the United States is 21 years (66).

We found no compelling evidence that behaviors that have been speculated to contribute to alcohol use (e.g., executive function, negative urgency, and impulsivity) are associated with prefrontal or insula GMV, leaving the behavioral mechanisms through which these GMVs may influence alcohol use unclear. DLPFC GMV was negatively correlated with delay discounting and using alcohol to cope with stress in our young adult sample (DNS sample) at nominal levels of significance, while these behavior characteristics and DLPFC GMV were unlinked in our young and middle-age adult sample (Tables S9,

Table 2. TWAS Discovery and Replication

Gene	CHR	Discovery—GTEx Frontal BA 9				Replication—CMC DLPFC			
		Locus Start	Locus End	TWAS z	TWAS p	Locus Start	Locus End	TWAS z	TWAS p
<i>C16orf93</i>	16	30772519	30772656	5.0152	5.30×10^{-7}	—	—	—	—
<i>CWF19L1</i>	10	102000000	102000000	−4.1674	3.08×10^{-5}	102000000	102000000	−2.11116	.0348
<i>PHBP9</i>	10	102000000	102000000	−3.8862	1.02×10^{-4}	—	—	—	—
<i>C18orf8</i>	18	21083473	21110576	3.831	1.28×10^{-4}	21083433	21111771	2.1613	.0307

Summary of transcriptome-wide association study (TWAS) results in the frontal cortex, conducted with FUSION. Discovery analysis: UK Biobank genome-wide association study ($N = 112,117$) and Genotype-Tissue Expression (GTEx) project gene expression ($N = 92$). Replication analysis: Alcohol Genome-Wide Consortium and the Cohorts for Heart and Aging Research in Genomic Epidemiology Plus consortia (AlcGen/CHARGE+) genome-wide association study ($N = 70,460$), and CommonMind Consortium (CMC) gene expression ($N = 452$). Empty rows in replication data indicate that the gene was not present in replication dataset. The p values are all uncorrected—all associations listed survive false discovery rate correction. Expression quantitative trait loci are available in the Supplemental Data.

BA, Brodmann area; CHR, chromosome; DLPFC, dorsolateral prefrontal cortex.

S10 in Supplement 1). This finding suggests that these behavioral factors may represent mechanisms through which GMV influences alcohol use in adolescence and young adulthood, potentially contributing to continued use, while GMV is uncorrelated with these behaviors as measured in later life. Nonetheless, it is also plausible that these nominally significant findings represent false-positives.

While both the discovery analysis in the DNS sample and the replication analysis in the HCP sample showed that alcohol use was significantly correlated with reduced GMV, the voxels of strongest association only partially overlap. Post hoc analyses found that the effect sizes of the association with atlas-defined ROIs were nearly identical in the 2 samples (Figure S2 in Supplement 1), further supporting the interpretation that identified GMV correlations with alcohol use are replicable. The limited overlap of peaks between the samples likely reflects the lower power in the HCP sample, which is a result of its smaller sample size and the family structure of the data, which resulted in even fewer independent observations. Several GMV findings in the DNS did not replicate, an outcome that may be attributable to differences between the samples (e.g., age), though the possibility that they are false-positives, or that null findings in the HCP sample are false-negatives, cannot be ruled out.

Substantiating the idea that it is biologically plausible that reduced GMV in the DLPFC represents a preexisting genetic liability for drinking, genomic risk for alcohol use was enriched only within brain gene sets. BA 9, which overlaps with the DLPFC regions identified in our neuroimaging analyses, was among the regions of strongest enrichment (Figure 4). Further, TWAS analyses revealed replicable evidence that genomic risk for alcohol use is associated with differential expression of *CWF19L1* and *C18orf8* within BA 9. While the function of these genes is not understood, both have been previously implicated in psychopathology and related traits, including schizophrenia, substance use, and cognition (Supplement 1), with rare mutations in *CWF19L1* causing autosomal recessive cerebellar ataxia (66,67), which is characterized by a loss of control of bodily movements, as well as developmental delay and mental retardation. Additional discussion of these findings and their limitations is presented in Supplement 1.

Given evidence that genetic liability is shared across substance use involvement (67) and other forms of psychopathology (68), our findings may generalize to other substances and overall psychopathology risk. While enrichment analyses implicate only brain pathways and TWASs identify replicable associations between genetic risk for alcohol consumption and gene expression in the frontal cortex, we cannot rule out the possibility that our observed effects are partially mediated by altered functioning of other pathways, such as alcohol metabolism in the liver (69). Moreover, the present results do not distinguish between reduced GMV as part of the mechanism by which genetic risk affects drinking behavior (10,70) and a pleiotropic effect of genetic risk on multiple outcomes (20).

We must note assumptions of heritability analyses including random mating and equal environments (71). On one hand, violations of the random-mating assumption would result in downwardly biased estimates of heritability (72). On the

other hand, violations of the equal environment assumption would result in upwardly biased estimates of heritability (72), though there is evidence that this bias, when present, is modest (73,74). An additional limitation of heritability analyses is that the statistical power required to parse the role of overlapping genetic and shared environmental factors is substantial, and it is beyond the scope of the current analysis. However, given that we found little evidence that shared environment contributes to the correlation of GMV and alcohol use, shared environment is an unlikely confound.

While our study is limited by our sample size, particularly of discordant siblings and longitudinal analyses, a major strength of our results is the convergent evidence provided by the different study designs (75). We note that our cross-sectional analyses of alcohol consumption and longitudinal analyses of adolescent use initiation are the largest to date that we know of. A primary limitation of our gene-expression analyses is that both of the gene-expression datasets included alcohol-exposed donors. Given the wide prevalence of alcohol use across the world (76), it will likely be impossible to ever definitively confirm in human adults that alcohol use is not confounding these results. Notably, none of the identified genes have been found to be differentially expressed in the frontal cortex of donors with alcoholism (77,78). Our analyses are also limited by the omission of the insula from the gene-expression data, precluding a comparison of the gene-expression correlates between the insula and frontal cortex.

Limitations notwithstanding, our study provides convergent evidence that smaller GMV in the insula and DLPFC associated with alcohol use may represent a genetically conferred liability that promotes early alcohol use. While early alcohol use may in turn lead to accelerated volume loss within these and other regions, these findings challenge predominant interpretations that smaller brain volumes tied to alcohol use emerge primarily from the atrophy-inducing effects of alcohol. As larger prospective samples are acquired (e.g., via the Adolescent Brain Cognitive Development study) (79), it will be interesting to examine the interplay of genetic risk and substance use on the trajectories of brain development.

ACKNOWLEDGMENTS AND DISCLOSURES

Data for this study were provided by the Human Connectome Project, WU-Minn Consortium (Grant No. 1U54MH091657; principal investigators David Van Essen, Ph.D., and Kamil Ugurbil, Ph.D.), which was funded by the 16 National Institutes of Health (NIH) institutes and centers that support the NIH Blueprint for Neuroscience Research and the McDonnell Center for Systems Neuroscience at Washington University. The Duke Neurogenetics Study was supported by Duke University and the National Institute on Drug Abuse (Grant No. DA033369 [to ARH]). The Teen Alcohol Outcomes Study (TAOS) was supported by Duke University and the National Institute on Alcohol Abuse and Alcoholism (Grant No. AA016274 [to DEW]). DAAB was supported by the NIH (Grant No. T32-GM008151) and the National Science Foundation (Grant No. DGE-1143954). LRF was supported by the National Institute on Alcohol Abuse and Alcoholism (Grant No. AA023693). CHD was supported by the NIH (Grant Nos. T32-DA007313 and T32-GM081739). AA was supported by the National Institute on Drug Abuse (Grant No. 5K02DA32573). ARH received additional support from the National Institute on Drug Abuse (Grant No. DA031579) and the National Institute on Aging (Grant No. AG049789). LMS was supported by the NIH (Grant Nos. K23

Alcohol Consumption and Brain Volume

AA025399 and U01 DA041093). RB was supported by the Klingenstein Third Generation Research and the NIH (Grant Nos. R01-AG045231, R01-HD083614, and R01-AG052564).

This article was published as a preprint on bioRxiv: <https://doi.org/10.1101/299149>.

All data pertaining to this study are available on request to the corresponding author. Datasets may also be accessed at the locations listed:

Duke Neurogenetics Study (DNS): <https://www.haririlab.com/projects/procedures.html>

Human Connectome Project (HCP): <https://www.humanconnectome.org/>

Teen Alcohol Onset Study (TAOS): Requests for data access should be submitted to the study principal investigator, Douglas Williamson, Ph.D.: douglas.williamson@duke.edu

UK Biobank: <http://www.ukbiobank.ac.uk>

Alcohol Genome-Wide Consortium and the Cohorts for Heart and Aging Research in Genomic Epidemiology Plus consortia (AlcGen/CHARGE+): Requests for access to summary statistics should be submitted to the study principal investigator, Gunter Schumann, Ph.D.: gunter.schumann@kcl.ac.uk

Genotype-Tissue Expression (GTEx)/CommonMind Consortium: Pre-computed gene expression weights were provided by the Gusev lab: <http://gusevlab.org/projects/fusion>; <https://gtexportal.org>; <https://www.nlm.nih.gov/genetics/commonmind>.

All authors report no biomedical financial interests or potential conflicts of interest.

ARTICLE INFORMATION

From the Department of Psychological and Brain Sciences (DAAB, CHD, AD, DMB, RB) and Department of Psychiatry (LRF, AA, ACH, DMB), Washington University, St. Louis, Missouri; Department of Psychiatry and Behavioral Sciences (NME, DEW) and Department of Psychology and Neuroscience (ARK, SRR, ARH), Duke University, and Durham VA Medical Center (DEW), Durham, North Carolina; and Department of Psychiatry and Behavioral Sciences (LMS), Medical University of South Carolina, Charleston, South Carolina.

Address correspondence to David Baranger, Ph.D., CB 1125 Psychological and Brain Sciences Building, Room 453, Washington University in St. Louis, One Brookings Drive, St. Louis, MO 63130; E-mail: dbaranger@gmail.com; and Ryan Bogdan, Ph.D., CB 1125 Psychological and Brain Sciences Building, Room 453, Washington University in St. Louis, One Brookings Drive, St. Louis, MO 63130; E-mail: rbogdan@wustl.edu.

Received May 13, 2019; revised Aug 19, 2019; accepted Aug 27, 2019.

Supplementary material cited in this article is available online at <https://doi.org/10.1016/j.biopsych.2019.08.029>.

REFERENCES

- World Health Organization (2014): Global status report on alcohol and health. Geneva, Switzerland: World Health Organization Press.
- Grant BF, Goldstein RB, Saha TD, Chou SP, Jung J, Zhang H, *et al.* (2015): Epidemiology of DSM-5 alcohol use disorder results from the National Epidemiologic Survey on Alcohol and Related Conditions III. *JAMA Psychiatry* 72:757–766.
- Substance Abuse and Mental Health Services Administration (2018): Key Substance Use and Mental Health Indicators in the United States: Results from the 2017 National Survey on Drug Use and Health. Rockville, MD: Center for Behavioral Health Statistics and Quality, Substance Abuse and Mental Health Services Administration. Available at: <https://www.samhsa.gov/data/sites/default/files/cbhsq-reports/NSDUHF2017/NSDUHF2017.pdf>. Accessed February 22, 2018.
- Mackey S, Allgaier N, Chaarani B, Spechler P, Orr C, Bunn J, *et al.* (2018): Mega-analysis of gray matter volume in substance dependence: general and substance-specific regional effects. *Am J Psychiatry* 17:19–128.
- Lange EHH, Nerland S, Jørgensen KNN, Mørch-Johnsen L, Nesvåg R, Hartberg CBB, *et al.* (2017): Alcohol use is associated with thinner cerebral cortex and larger ventricles in schizophrenia, bipolar disorder and healthy controls. *Psychol Med* 4:55–668.
- Yang X, Tian F, Zhang H, Zeng J, Chen T, Wang S, *et al.* (2016): Cortical and subcortical gray matter shrinkage in alcohol-use disorders: A voxel-based meta-analysis. *Neurosci Biobehav Rev* 66:92–103.
- Thayer RE, YorkWilliams S, Karoly HC, Sabbineni A, Ewing SF, Bryan AD, Hutchison KE (2017): Structural neuroimaging correlates of alcohol and cannabis use in adolescents and adults. *Addiction* 112:2144–2154.
- Whelan R, Watts R, Orr CA, Althoff RR, Artiges E, Banaschewski T, *et al.* (2014): Neuropsychosocial profiles of current and future adolescent alcohol misusers. *Nature* 512:185–189.
- Pfefferbaum A, Kwon D, Brumback T, Thompson WK, Cummins K, Tapert SF, *et al.* (2017): Altered brain developmental trajectories in adolescents after initiating drinking. *Am J Psychiatry* 175:370–380.
- Holmes AJ, Hollinshead MO, Roffman JL, Smoller JW, Buckner RL (2016): Individual differences in cognitive control circuit anatomy link sensation seeking, impulsivity, and substance use. *J Neurosci* 36:4038–4049.
- Taffe MA, Kotzebue RW, Crean RD, Crawford EF, Edwards S, Mandyam CD (2010): Long-lasting reduction in hippocampal neurogenesis by alcohol consumption in adolescent nonhuman primates. *Proc Natl Acad Sci U S A* 107:11104–11109.
- Kühn S, Gallinat J (2013): Gray matter correlates of posttraumatic stress disorder: A quantitative meta-analysis. *Biol Psychiatry* 73:70–74.
- Shnitko TA, Liu Z, Wang X, Grant KA, Christopher D (2019): Chronic alcohol drinking slows brain development in adolescent and young adult nonhuman primates. *eNeuro* 6:1–11.
- Zou X, Durazzo TC, Meyerhoff DJ (2018): Regional brain volume changes in alcohol-dependent individuals during short-term and long-term abstinence. *Alcohol Clin Exp Res* 42:1062–1072.
- Luciana M, Collins PF, Muetzel RL, Lim KO (2013): Effects of alcohol use initiation on brain structure in typically developing adolescents. *Am J Drug Alcohol Abuse* 39:345–355.
- Squeglia LM, Tapert SF, Sullivan EV, Jacobus J, Meloy MJ, Rohlfing T, Pfefferbaum A (2015): Brain development in heavy-drinking adolescents. *Am J Psychiatry* 172:531–542.
- Seo S, Beck A, Matthis C, Genauck A, Banaschewski T, Bokde ALW, *et al.* (2018): Risk profiles for heavy drinking in adolescence: Differential effects of gender. *Addict Biol* 21:348–356.
- Windle M, Gray JC, Mankit K, Barton AW, Brody G, Beach SRH, *et al.* (2018): Age sensitive associations of adolescent substance use with amygdalar, ventral striatum, and frontal volumes in young adulthood. *Drug Alcohol Depend* 186:94–101.
- Squeglia LM, Gray KM (2016): Alcohol and drug use and the developing brain. *Curr Psychiatry Rep* 18:46.
- Dager AD, McKay DR, Kent JW, Curran JE, Knowles E, Sprooten E, *et al.* (2015): Shared genetic factors influence amygdala volumes and risk for alcoholism. *Neuropsychopharmacology* 40:412–420.
- Henderson KE, Vaidya JG, Kramer JR, Kuperman S, Langbehn DR, O'Leary DS (2018): Cortical thickness in adolescents with a family history of alcohol use disorder. *Alcohol Clin Exp Res* 42:89–99.
- Wilson S, Malone SM, Thomas KM, Iacono WG (2015): Adolescent drinking and brain morphometry: A co-twin control analysis. *Dev Cogn Neurosci* 16:130–138.
- Sharma VK, Hill SY (2017): Differentiating the effects of familial risk for alcohol dependence and prenatal exposure to alcohol on offspring brain morphology. *Alcohol Clin Exp Res* 41:312–322.
- Van Essen DC, Smith SM, Barch DM, Behrens TEJ, Yacoub E, Ugurbil K (2013): The WU-Minn Human Connectome Project: An overview. *Neuroimage* 80:62–79.
- Swartz JR, Williamson DE, Hariri AR (2015): Developmental change in amygdala reactivity during adolescence: Effects of family history of depression and stressful life events. *Am J Psychiatry* 172:276–283.
- Nikolova YS, Knodt AR, Radtke SR, Hariri AR (2016): Divergent responses of the amygdala and ventral striatum predict stress-related problem drinking in young adults: possible differential markers of

- affective and impulsive pathways of risk for alcohol use disorder. *Mol Psychiatry* 21:348–356.
27. Gusev A, Ko A, Shi H, Bhatia G, Chung W, Penninx BWJH, *et al.* (2016): Integrative approaches for large-scale transcriptome-wide association studies. *Nat Genet* 48:245–252.
 28. Clarke T-K, Adams MJ, Davies G, Howard DM, Hall LS, Padmanabhan S, *et al.* (2017): Genome-wide association study of alcohol consumption and genetic overlap with other health-related traits in UK Biobank ($N = 112\,117$). *Mol Psychiatry* 22:1376–1384.
 29. Schumann G, Liu C, O'Reilly P, Gao H, Song P, Xu B, *et al.* (2016): KLB is associated with alcohol drinking, and its gene product β -Klotho is necessary for FGF21 regulation of alcohol preference. *Proc Natl Acad Sci U S A* 113:14372–14377.
 30. The GTEx Consortium, Welter D, MacArthur J, Morales J, Burdett T, Hall P, *et al.* (2015): The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans. *Science* 348:648–660.
 31. Fromer M, Roussos P, Sieberts SK, Johnson JS, Kavanagh DH, Perumal TM, *et al.* (2016): Gene expression elucidates functional impact of polygenic risk for schizophrenia. *Nat Neurosci* 19:1442–1453.
 32. Saunders JB, Aasland OG, Babor TF, de la Fuente JR, Grant M (1993): Development of the Alcohol Use Disorders Identification Test (AUDIT): WHO Collaborative Project on Early Detection of Persons With Harmful Alcohol Consumption—II. *Addiction* 88:791–804.
 33. Babor TF, Higgins-Biddle JC, Saunders JB, Monteiro MG (2001): The Alcohol Use Disorders Identification Test: Guidelines for Use in Primary Care. Geneva, Switzerland: World Health Organization.
 34. Bucholz KK, Cadoret R, Cloninger CR, Dinwiddie SH, Hesselbrock VM, Nurnberger Jr JI, *et al.* (1994): A new, semi-structured psychiatric interview for use in genetic linkage studies: A report on the reliability of the SSAGA. *J Stud Alcohol* 55:149–158.
 35. Molina BSG, Flory K, Hinshaw SP, Greiner AR, Arnold LE, Swanson JM, *et al.* (2007): Delinquent behavior and emerging substance use in the MTA at 36 months: Prevalence, course, and treatment effects. *J Am Acad Child Adolesc Psychiatry* 46:1028–1040.
 36. Kendler KS, Gardner CO, Hickman M, Heron J, Macleod J, Lewis G, Dick DM (2014): Socioeconomic status and alcohol-related behaviors in mid- to late adolescence in the Avon Longitudinal Study of Parents and Children. *J Stud Alcohol Drugs* 75:541–545.
 37. Meng Y, Holmes J, Hill-McManus D, Brennan A, Meier PS (2014): Trend analysis and modelling of gender-specific age, period and birth cohort effects on alcohol abstinence and consumption level for drinkers in Great Britain using the General Lifestyle Survey 1984–2009. *Addiction* 109:206–215.
 38. Collins SE (2016): Associations between socioeconomic factors and alcohol outcomes. *Alcohol Res* 38:83–94.
 39. Grittner U, Kuntsche S, Gmel G, Bloomfield K (2013): Alcohol consumption and social inequality at the individual and country levels—Results from an international study. *Eur J Public Health* 23:332–339.
 40. Delker E, Brown Q, Hasin DS (2016): Alcohol consumption in demographic subpopulations: An epidemiologic overview. *Alcohol Res* 38:7–15.
 41. Cacciola EET, Nevid JS (2014): Alcohol consumption in relation to residence status and ethnicity in college students. *Psychol Addict Behav* 28:1278–1283.
 42. Keyes KM, Hatzenbuehler ML, Grant BF, Hasin DS (2012): Stress and alcohol: Epidemiologic evidence. *Alcohol Res* 34:391–400.
 43. Enoch M-A (2011): The role of early life stress as a predictor for alcohol and drug dependence. *Psychopharmacology (Berl)* 214:17–31.
 44. McLaughlin KA, Green JG, Gruber MJ, Sampson NA, Zaslavsky AM, Kessler RC (2013): Childhood adversities and adult psychiatric disorders in the National Comorbidity Survey Replication II. 67:124–132.
 45. Tzourio-Mazoyer N, Landeau B, Papathanassiou D, Crivello F, Etard O, Delcroix N, *et al.* (2002): Automated anatomical labeling of activations in SPM using a macroscopic anatomical parcellation of the MNI MRI single-subject brain. *Neuroimage* 15:273–289.
 46. Winkler AM, Webster MA, Vidaurre D, Nichols TE, Smith SM (2015): Multi-level block permutation. *Neuroimage* 123:253–268.
 47. Winkler AM, Ridgway GR, Webster MA, Smith SM, Nichols TE (2014): Permutation inference for the general linear model. *Neuroimage* 92:381–397.
 48. Winkler AM, Ridgway GR, Douaud G, Nichols TE, Smith SM (2016): Faster permutation inference in brain imaging. *Neuroimage* 141:502–516.
 49. Vul E, Harris C, Winkelman P, Pashler H (2009): Puzzlingly High Correlations in fMRI Studies of Emotion, Personality, and Social Cognition1. *Perspect Psychol Sci* 4:274–290.
 50. Kochunov P, Jahanshad N, Marcus D, Winkler A, Sprooten E, Nichols TE, *et al.* (2015): Heritability of fractional anisotropy in human white matter: A comparison of Human Connectome Project and ENIGMA-DTI data. *Neuroimage* 111:300–301.
 51. Ziyatdinov A, Brunel H, Martinez-Perez A, Buil A, Perera A, Soria JM (2016): Solaris: An R interface to SOLAR for variance component analysis in pedigrees. *Bioinformatics* 32:1901–1902.
 52. Revelle W (2019): Package 'psych': Procedures for Psychological, Psychometric and Personality Research. Version 1.8.10. Available at: <https://cran.r-project.org/package=psych>.
 53. Bates D, Maechler Martin, Walker S (2019): Package 'lme4': Linear Mixed-Effects Models Using 'Eigen' and S4. Version 1.1-19. Available at: <https://cran.r-project.org/package=lme4>.
 54. R Core Team (2014): R: A language and environment for statistical computing. Vienna, Austria: R Foundation for Statistical Computing. Version 3.5.1. Available at: <http://www.r-project.org/>.
 55. Hahsler M, Piekenbrock M, Arya S, Mount D (2017): dbscan: Density Based Clustering of Applications with Noise (DBSCAN) and Related Algorithms. Version 1.1-3. Available at: <https://cran.r-project.org/package=dbscan>.
 56. Pinheiro J, DebRoy S, Bates D, Sarkar D, R Core Team (2017): nlme: Linear and Nonlinear Mixed Effects Models. Version 3.1-137. Available at: <https://cran.r-project.org/package=nlme>.
 57. Keller MC (2014): Gene \times environment interaction studies have not properly controlled for potential confounders: The problem and the (simple) solution. *Biol Psychiatry* 75:18–24.
 58. Baranger DAA, Ifrah C, Prather AA, Carey CE, Corral-Frías NS, Drabant Conley E, *et al.* (2016): PER1 rs3027172 genotype interacts with early life stress to predict problematic alcohol use, but not reward-related ventral striatum activity. *Front Psychol* 7:464.
 59. Bates D, Mächler M, Bolker BM, Walker SC, Maechler Martin, Walker SC (2015): Fitting linear mixed-effects models using lme4. *J Stat Softw* 67:1–48.
 60. Finucane HK, Bulik-Sullivan B, Gusev A, Trynka G, Reshef Y, Loh PR, *et al.* (2015): Partitioning heritability by functional annotation using genome-wide association summary statistics. *Nat Genet* 47:1228–1235.
 61. Finucane HK, Reshef Y, Anttila V, Slowikowski K, Gusev A, Byrnes A, *et al.* (2018): Heritability enrichment of specifically expressed genes identifies disease-relevant tissues and cell types. *Nat Genet* 50:621–629.
 62. Bulik-Sullivan B, Loh PR, Finucane HK, Ripke S, Yang J, Patterson N, *et al.* (2015): LD score regression distinguishes confounding from polygenicity in genome-wide association studies. *Nat Genet* 47:291–295.
 63. de Leeuw CA, Mooij JM, Heskes T, Posthuma D (2015): MAGMA: Generalized gene-set analysis of GWAS data. *PLoS Comput Biol* 11: e1004219.
 64. Watanabe K, Taskesen E, Van Bochoven A, Posthuma D (2017): Functional mapping and annotation of genetic associations with FUMA. *Nat Commun* 8:1826.
 65. Watanabe K, Stringer S, Frei O, Umičević Mirkov M, de Leeuw C, Polderman TJC, *et al.* (2018): A global overview of pleiotropy and genetic architecture in complex traits. *Nat Genet* 51:1339–1348.
 66. Young-Wolff KC, Enoch MA, Prescott CA (2011): The influence of gene-environment interactions on alcohol consumption and alcohol use disorders: A comprehensive review. *Clin Psychol Rev* 31:800–816.

Alcohol Consumption and Brain Volume

67. Carey CE, Agrawal A, Bucholz KK, Hartz SM, Lynskey MT, Nelson EC, *et al.* (2016): Associations between polygenic risk for psychiatric disorders and substance involvement. *Front Genet* 7:149.
68. Smoller JW, Craddock N, Kendler K, Lee PH, Neale BM, Nurnberger Jr, *et al.* (2013): Identification of risk loci with shared effects on five major psychiatric disorders: a genome-wide analysis. *Lancet* 381:1371–1379.
69. Dick DM, Agrawal A (2008): The genetics of alcohol and other drug dependence. *Alcohol Res Health* 31:111–118.
70. Drouman V, Read SJ, Bechara A (2015): Revisiting the role of the insula in addiction. *Trends Cogn Sci* 19:414–420.
71. Rijdsdijk FV, Sham PC (2002): Analytic approaches to twin data using structural equation models. *Brief Bioinform* 3:119–133.
72. Agrawal A, Lynskey MT (2008): Are there genetic influences on addiction: Evidence from family, adoption and twin studies. *Addiction* 103:1069–1081.
73. LaBuda MC, Svikiel DS, Pickens RW (1997): Twin closeness and co-twin risk for substance use disorders: Assessing the impact of the equal environment assumption. *Psychiatry Res* 70:155–164.
74. Felson J (2014): What can we learn from twin studies? A comprehensive evaluation of the equal environments assumption. *Soc Sci Res* 43:184–199.
75. Munafò MR, Davey Smith G (2018): Robust research needs many lines of evidence. *Nature* 553:399–401.
76. Substance Abuse and Mental Health Services Administration (2015): Results from the 2015 National Survey on Drug Use and Health: Detailed Tables. Available at: <https://www.samhsa.gov/data/report/results-2015-national-survey-drug-use-and-health-detailed-tables>. Accessed February 22, 2018.
77. Liu J, Lewohl JM, Harris RA, Iyer VR, Dodd PR, Randall PK, Mayfield RD (2006): Patterns of gene expression in the frontal cortex discriminate alcoholic from nonalcoholic individuals. *Neuropsychopharmacology* 31:1574–1582.
78. Kapoor M, Wang J, Farris SP, Liu Y, McClintick J, Gupta I, *et al.* (2019): Analysis of whole genome-transcriptomic organization in brain to identify genes associated with alcoholism. *Transl Psychiatry* 9:89.
79. Volkow ND, Koob GF, Croyle RT, Bianchi DW, Gordon JA, Koroshetz WJ, *et al.* (2018): The conception of the ABCD study: From substance use to a broad NIH collaboration. *Dev Cogn Neurosci* 32:4–7.